# Antimicrobial And Phytochemical Potential Of Chenopodium Album Linn.

## Rashi Saini, Dinesh Kumar, Amita Mittal

Abstract: Chenopodium album Linn. (Family: Chenopodiaceae) is a wildly growing weed in Africa, Asia, Europe and North America. In folk medicine, this plant has been used as antiparasitic, diuretic, hepato-protective, laxative and sedative. Its leaves possesses antirheumatic, anthelmentic, antiphlogistic, mildly laxative and odontalogic properties, applied as wash or poultice to bug bites, rheumatic joints, sunstroke, and swollen feets. This plant is used as vegetable in India and has high biological potential in addition to basic nutritional benefits. Previous scientific studies have reported the anthelmintic, antipruritic, contraceptive and sperm immobilizing properties of this plant. Hence, this plant possesses a great potential for detailed biological screening. So, in order to elucidate the potential traditional use of this plant, the present study was undertaken to evaluate its phytochemical and antimicrobial screening along with its free radical scavenging effect. Phytochemical investigations confirmed the presence of alkaloids, flavonoids, anthocynidine, saponins, glycosides, tannins and carbohydrates in the plant leaves extract. Antimicrobial effect tested by well diffusion method proved that methanolic leaf extract of the plant possesses considerable inhibitory activity against Escherichia coli, Pseudomonas aeruginosa, Bacillus subtilis, Candida albicans, and Candida glabrata. The antioxidant activity was studied by DPPH free radical scavenging activity, total phenolics content and ascorbic acid estimation. The decolorization of purple colour in DPPH assay indicated the free radical scavenging activity. This activity was dose dependent and maximum activity (80.58%) was observed 50 µg/ ml. The total phenolic content in C. album is found to be 0.115g of phenols, and is calculated by plotting O.D. on standard graph of catechol. The ascorbic acid in leaf extract of C. album is calculated as 1.29 mg of ascorbate / g of leaf. Total phenolic content and ascorbic acid in the leaf extract justify the nutritiona

Index Terms: Phytochemical, antioxidant, antimicrobial, P. aeruginosa, C.album.

# 1 INTRODUCTION

HERBS are still used by 80% of world population, because of their easy availability and very negligible side effects. From ancient period medicinal plants are used for pharmacological studies. Knowledge of therapeutic value of these plants acts as sources for lead compound development [1]. Chenopodium album Linn. (C. album) belonging to family Chenopodiaceae, is commonlycalled as Bathua(Hindi), Chandanbetu (Bengali), Parupukkirai (Tamil), Pappukura (Telgu) Katuayamoddakam (Malyalam) in India. It is afast growing annual plantwhich wildlygrows as weed in Africa, Asia, Europe and North America. Tender branches and leaves are used as a vegetable in various parts of world. In India, it is locally used in curd known as "Raita". Leaves are often dried and stored for later use. Infusions of leaves were also used as a laxative. Hindu physicians recommended C. album for treatment of splenic enlargement and liver diseases [2]. In folk medicine, this plant has been used as antiparasitic, diuretic, hepatoprotective, laxative and sedative. Its leaves possesses antirheumatic, anthelmentic, antiphlogistic, mildly laxative and odontalgic properties, applied as wash or poultice to bug bites, rheumatic joints, sunstroke, and swollen feet [3].

This plant is used as vegetable in India and has high biological potential in addition to basic nutritional benefits. Previous scientific studies have reported the anthelmintic, antipruritic, antinociceptive, contraceptive and sperm immobilizing properties of this plant [4]. Hence, this plant possesses a great potential for detailed biological screening. So, in order to elucidate the potential traditional use of this plant, the present study was undertaken to evaluate C. album leaf extracts for phytochemical and antimicrobial screening along with its free radical scavenging effect.

## **2 MATERIALS AND METHOD**

## 2.1 Collection of plant material

C. album leaves were taken from village Surkhpur (Kurukshetra, Haryana) in December 2018. After drying in shade, the leaves were coarsely powdered.

#### 2.2 Preparation of the plant extracts:

For extract preparation, the leaf powder was soaked in various organic solvents (methanol, chloroform, n-hexane, petroleum ether and acetone) in the ratio of 1:10 (20gm in 200ml solvent) for 72 hr. at 30°C. After maceration of 72 hr, extracts were filtered. Complete evaporation of solvent was done in water bath according to the boiling temperature of respective solvent. In the residual powder 5% (w/v) DMSO was added and stored at 4°C till further investigation.

# 2.3 Study of the Antibacterial Activity

## Culture collection

The microbial strains were obtained from MTCC Centre, IMTECH, Chandigarh. The pure cultures were preserving on nutrient agar for growth. Cultures were stored at 4°C in refrigerator for further use.

## Preparation of extract dilutions

1 mg ml<sup>-1</sup> extract concentration was prepared by adding 10 mg of extract in 10 ml of 5% dimethylsulfoxide, and further diluted to 0.5 mg/ml with DMSO.

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#### Agar well diffusion method

In this method, lawn culture of bacteria (B. subtilis, E. coli, P. aeruginosa) and fungi (C. albicans and C. glabrata) were spread on the Muller Hinton agar using spreader. The wells were taken from agar plates using a cork borer. The extracts were poured into the well. The plates were placed in BOD incubator at 37°C for 24 hrs. After incubation, the zone of inhibition was determined [5].

## 2.4 Phytochemical Screening

Prepared C. album extracts were screened for various biochemical parameters [6].

## 2.5 Antioxidant activity

## DPPH radical scavenging method

In this method, solution of 0.1 mM DPPH was prepared in methanol and from this solution 1ml was mixed with 3 ml of extract solution prepared in methanol having 10, 20, 30, 40 and 50  $\mu$ g/ml of leaf extract. Reaction mixture was swirled thoroughly. After 30 minute incubation, absorbance of the mixture was observed at 517 nm in Double beam UV-Visible spectrophotometer [7]. L-ascorbic acid was taken as standard. The scavenging ability of sample was estimated as per reported method [8].

% Inhibition = [(Ac-Aa)/Ac] X 100

Where Ac and Aa represent the absorption of the control and extract respectively.

# Estimation of total phenols

500 mg leaves of C. album were mixed with 80% ethanol (10

times) followed by centrifugation at 10,000 rpm for half an hour. Supernatants were mixed together and evaporated to dryness. Again, remains were dissolved in a known quantity of distilled water and transferred into test tubes by pipette. Final volume was made 3.0 ml with distilled water. 0.5 ml of Folin-Phenol reagent was added to each test tube and mixed well. 3 minute later, 2.0 ml of 20% sodium carbonate solution was added. After mixing, tubes were placed in boiling water for 1 minute and then cooled. Absorbance was noted at 650 nm in UV-Vis spectrophotometer against blank [9].

#### Ascorbic acid Estimation

Ascorbic acid content existing in C. album leaves was determined as per the previous method [10] [11].

#### **3 RESULTS AND DISCUSSION**

## 3.1 Antimicrobial activity:

The antimicrobial potential of different leaf extracts of C. album is mentioned in Table 1. The results shows inhibitory effect of extract on growth of microbial species: E.coli, P. aeruginosa (Gram-ve), B. subtilis (Gram +ve) and C. albicans, C. glabrata (Fungi). The methanolic leaf extract showed maximum activity against all strains with highest inhibition zone of 31 mm against gram positive B. subtilis, while the chloroform and acetone extracts were also active against test samples with active range from 14-28 mm and 11-16 mm respectively. n-Hexane extract had no effect on any microbial species except E. coli and C. albican with 10 mm inhibition zone, while petroleum ether extract showed no activity. Highest antimicrobial activity was also observed by Külcü et al in C. album [12]. Ramproshad et al. also reported great antimicrobial activity against various pathogenic microbes [13].

**Table 1:** Antimicrobial potential of C. album leaves extracts.

		Diameter of inhibition zone (mm)										
Micro	Methanol		Chloroform		Acetone		n-hexane		Pet. Ether			
organisms	0.5	1	0.5	1	0.5	1	0.5	1	0.5	1		
	mgml <sup>-1</sup>	mgml⁻¹	mgml <sup>-1</sup>	mgml <sup>-1</sup>	mgml <sup>-1</sup>	mgml⁻¹	mgml <sup>-1</sup>	mgml⁻¹	mgml⁻¹	mgml <sup>-1</sup>		
E. coli	14	25	14	21	11	15	10					
P. aeruginosa	18	24	20	23	12	16						
B. subtilis	15	31	25	28	12							
C. albicans	17	21	16	20	12	15	10					
C. glaburata	18	24	19	23	11	15						

## 3.2 Phytochemical analysis:

Main phytoconstituents such as flavonoids, tannins, carbohydrates, proteins, phenolic compounds, alkaloids and sterols were found to be present in the aqueous and methanolic leaf extracts (table 2). Results are in accordance with the former studies [14]. Result shows the presence of alkaloids, phenolic, flavonoids, saponin, tannin, protein, carbohydrates, and glycoside, in the leaf extract of C. album. It

has been already reported that alkaloids are potent anesthetic and spasmolytic agents whereas saponins improves the immunity [15] [16]. The finding of the study shows that this plant has a number of chemical constituents, which might be responsible for the different pharmacological activities [17]. Therefore, presence of phenolic compounds in the leaves could be responsible for their free radical scavenging activity.

Phytochemicals	Aqueous extract	Methanolic extract	
Alkaloids	-	+	
Amino acids	+	=	
Proteins	+	-	
Carbohydrates	+	+	
Flavonoids	+	+	
Anthraquinone	-	-	
Cardiac glycosides	+	-	
Starch	+	+	
Saponin	+	+	
Anthocynidine	+	+	
Steroids	+	+	
Tannins	+	+	
Phenolic compounds	+	+	

Concentration (μg/ml)	Absorbance at 517 nm		% Inhibition of c. album		
10	560		45.098		
20	428		58.039		
30	392		61.560		
40	224		78.039		
50	198		80.580		
Standard (control)	1020				

Table 3: DPPH scavenging activity of methanolic extract of C. album

Table 2: Phytochemical screening of aqueous and methanolic leaf extracts of C. album. ("+ve" indicates presence of compound where as "-ve" indicates absence of compound)

#### 3.3 Antioxidant assay

## **DPPH free radical scavenging activity:**

Scavenging activity of methanolic extract of C. album is shown in table 3. A significant drop in the concentration of DPPH free radicals was observed due to scavenging ability of extract. The activity was dose dependent. Maximum scavenging activity 80.580 was observed at 50 µg/ml concentration. The total antioxidant capacity in the leaf extract was determined by the formation of phosphomolybdenum complex, which was able to reduce the stable DPPH radical to the yellow colored diphenylpicrylhydrazine. Free radical scavenging activity of this plant has also been reported previously [18]. Lone et al. (2017) reported 45%-73% DPPH activity of methanol extract of C.album [19]. 9.28% to 24.90% activity was observed by Külcü et al [12].

# Total phenolics content:

Hydroxyl group of Phenolic compounds is responsible for scavenging ability of plant. It was observed by the formation of deep blue colour. The total phenolics content in C. album is 0.115g of phenols, calculated by plotting O.D. (Optical Density) on standard graph of catechol. Phytochemical studies demonstrated that C. album methanolic extract have flavanoids, alkaloids, tannins and to a very little extent of carbohydrates and glycosides etc. Flavonoids and phenolics or their combinations found to be liable for antioxidant potential of plant. The phyto constituents like polyphenol, flavonoid and isoflavonoid have gained considerable interest in improvement of human health, especially in tumor reduction. Phenolic compounds have been found significant due to their accepted role in the prevention of various human health problems [20]. Estimation of ascorbic acid (Vitamin C): A naturally present antioxidant in fruits, vegetables, medicinal plants and whole grains, plays a principal role in averting free radicals. The value was estimated by plotting the O.D. at 650 nm on standard graph of ascorbic acid and is calculated as 1.29 mg of ascorbate / g of leaf. Vitamin C being immunity booster has positive effect on our health and body.

## 4 CONCLUSIONS:

Owing to the frequent use of this plant in Indian cuisine and because of its health benefits, study was performed with the aim to explore antimicrobial, phytochemical and antioxidant potential. Resultof the study indicates that Chenopodium has potent antimicrobial potential and very good antioxidant effect. The activity of this plant might be due to the occurrence of one or more of the chemical constituents reported in extract. The activity may be useful in the treatment of various health problems and diseases.

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## 6 REFERENCES

- [1] P. Samanta, and D. Somnath, "Study on the Antibacterial, Antioxidant Activities and Phytochemical Analysis of Medicinal Plants in West-Bengal, India". International Journal of Current Microbiology and Applied Science. 4: pp. 116-123, 2015.
- [2] S. Priya, S. Yogesh, A. K. Singhai and S. Abhishek, "Pharmacological and phytochemical profile of C. album linn." Research Journal of Pharmacy and Technology. 3 pp. 960-966, 2010.
- [3] N. Z. Kokanova, P. Nedialkov and S. Nikolov, "The genus chenopodium: phytochemistry, ethnopharmacology and pharmacology". Pharmacognosy Review. 3: pp. 280-306, 2009.
- [4] A. Poonia and A. Upadhayay, "Chenopodium album Linn: review of nutritive value and biological properties". Journal of Food Science and Technology. 52(7): pp. 3977- 85, 2015.
- [5] A.W. Bauer, W. M. Kirby, J. C. Sheris and M. Turck, "Antibiotic susceptibility testing by a standardized single disc method". Journal of Clinical Pathology. 45: pp. 149-158, 1966.
- [6] G. E. Trease, and W. C. Evans, Pharmacognosy. (15th Ed.). London: Saunders Publishers, 2002.
- [7] Blois, "Antioxidant determination by the use of a stable free radical". Nature. 181. Pp. 1199-1200. 1958.
- [8] O. P. G. Nmorsi, N. C. D. Ukwandu and A. O. Egwunyenga, "Antioxidant status of Nigerian children

- with Plasmodium falciparum malaria". African Journal of Microbiology Research. 5: pp. 61- 64, 2007.
- [9] C. P. Malick, and M.B. Singh, New Delhi: Kalyani Publisher. Plant Enzymology and Histo Enzymology, 1980.
- [10] J. H. Roe and C. A. Kuether, "The Determination of Ascorbic acid in Whole blood and Urine Through 2, 4-Dinitrophenylhydrazine Derivative of Dehydro ascorbic acid". Journal of Biological Chemistry.147: pp. 399-.407, 1943.
- [11] S. Sadasivam, and B. Theyomoli, In: Practical Manual of Biochemistry, Tamilnadu Agriculture University, Coimbatore, 1987.
- [12] D.B. Külcü, C.D. Gökşk, and S. Aydn, "An Investigation of Antibacterial and Antioxidant Activity of Nettle (Urtica dioica L.), Mint (Mentha piperita), Thyme (Thyme serpyllum) and Chenopodium album L. Plants from Yaylack Plateau, Giresun, Turkey". Turkish Journal of Agriculture Food Science and Technology, 7(1): pp. 73-80, 2019
- [13] S. Ramproshad, B. Mondal, M. Mondal, S. J. Uddin, and M.G. Hossain, "Comparative Phytochemical & Pharmacological Study on Enhydra fluctuans, Alternanthera philoxeroides and Chenopodium album". Pharmacologyonline. 3 pp. 337- 353, 2018.
- [14] M. Kaur, S. Sharma, S. Garg and M. Arora, "Study of Antibacterial Activity of Chenopodium album Leaves Extract". International Journal of Pharmacy and Pharmaceutical Research. 10, pp. 1-4, 2018.
- [15] A. Adedapo, F. Jimoh, and A. Afolayan, "Comparison of the nutritive value and biological activities of the acetone, methanol and water extracts of the leaves of Bidens pilosa and Chenopodium album". Acta Poloniae Pharmaceutica. 68: 83-92, 2011.
- [16] K. Gupta, and D.S. Wagle, "Nutritional and anti nutritional factors of green leafy vegetables". Journal of Agricultural and Food Chemistry. 36: pp. 472-74, 1988.
- [17] M. Valko, D. Leibfritz, J. Moncol, M. T. Cronin, M. Mazur and J. Telser, "Free radicals and antioxidants in normal physiological functions and human disease". International Journal of Biochemistry and Cell Biology. 39, pp. 44-84, 2007.
- [18] S. Kumar and D. Kumar, "Antioxidant and Free Radical Scavenging Activity of Edible Weeds". African Journal of Food, Agriculture, Nutrients and Development. 9(5). Pp. 1174-1190, 2009.
- [19] B. A. Lone, M. Z. Chishti., F. A. Bhat, H. Tak, S. A. Bandh, and A. Chan, "Evaluation of anthelmintic, antimicrobial and antioxidant activity of Chenopodium album". Tropical Animal Health and Production, 49(8): pp. 1597-1605, 2017.
- [20] A. H. Laghari, S. Memona, A. Nelofar, K. M. Khan, and A. Yasmin, "Determination of free phenolic acids and antioxidant activity of methanolic extracts obtained from fruits and leaves of Chenopodium album". Food Chemistry. 126: pp. 1850-1855, 2011.